JCRT.ORG

ISSN: 2320-2882



INTERNATIONAL JOURNAL OF CREATIVE **RESEARCH THOUGHTS (IJCRT)**

An International Open Access, Peer-reviewed, Refereed Journal

BIOACTIVE COMPOUNDS FROM MANGROVE **BARK**

¹Powar Pratishtha S. and ²Gaikwad Dattatraya K.

¹Assistant professor, ²Professor; Regional Director, ¹Department of Botany, Dada Patil Mahavidyalaya Karjat, Dist. Ahmednagar, MS, India.

Abstract

The aim of the present study was carried out to determine the possible chemical compounds of methanolic extract of outer bark of Cynometra iripa (Fam. Leguminoceae) and Lumnitzera racemosa (Fam. Combetaceae). The phytochemical compound screened by GC-MS method from stem bark by using GCMS technique. The GCMS analysis revealed that the presence of bioactive compounds. The main constituents were 9-12 Octadecadienoic acid, 4-Methylmannose, 7-Tridecanone in higher Area % and , Sitosterol, Ergosta-8, 24(28)-dien-3ol, 4, 14-dimethyl, Glycerin, 2 Heptanone-3 methyl, α-L-Galactopyranoside, methyl6-deox, 3-O-Methyl-d-glucose in lower Area %. The bioactive compounds are found active against antimicrobial and anti-inflammatory activities.

Index Terms - Antimicrobial, Bark, Cymnometra, GCMS, Lumnitzera.

I.INTRODUCTION

In the tropics and subtropical regions of world, types of plant called, Mangroves are present. They contain variety of trees and shrubs that grow in saline coastal habitats mainly between latitudes 25° N and 25° S. The forests of mangroves occupy near about only 1% of area of the world (Saenger, 2002). Kathiresan and Bingham, (2001) described that mangrove have recognized in variety of plants distributed in 22 genera from 16 families and 65 different Mangrove species. Due to excess types of stresses such as high and low tide, extreme humidity and salinity found in Mangroves. To defend this type of conditions they produce high sources of bioactive compounds. Variety of secondary metabolites like alkaloids, tannins, terpenoids, phenolics, flavonoids, steroids etc. have been characterized from mangroves which have prime importance in pharmacological, toxicological, and ecological types (Kokpal et al., 1990 and Bandaranayake, 2002), they further reported that Mangrove sources are commonly traditionally useful in folklore medicines and The plant part extracts obtained from large species of mangrove have recognized activities against pathogens of plant, animals including human being. The outermost covering of stem found in woody plants is called as bark. These are the mass of tissues found outer side to the vascular cambium or it is present over the central bundle of vascular cylinder. Therefore, there is an imperative necessity to find out phytochemical compounds present in the bark of stem of Mangroves.

II. MATERIAL AND METHODS

The bark samples of mangrove bark of Cynometra iripa (Fam. Leguminosae) and Lumnitzera racemosa (Fam. Combretaceae) were collected from estuaries of Sindhudurga and Ratnagiri Districts in the month of December.

2.1 Preparation of powder and extract

Stem bark were shade dried, powdered and extracted with Methanol for 6-8 hours using Soxhlet apparatus. The extract was then filtered through muslin cloth, evaporated dried to get the viscous residue. The methanolic extracts of the plant was used for GC-MS analysis. 1 µl of the methanolic bark extract of root and stem was employed for GC-MS analysis.

2.2 GC MS Analysis

GC-MC Plays a key role in the analysis of unknown components of plant origin. The methanolic extract obtained from mangrove bark was subjected to Gas Chromatography and Mass Spectroscopy for the quantitative determination of phytochemicals extracted from mangrove bark. Some of the important features are summarized below.

III. Instrumentation

GC: Shimadzu Make QP-2010 with non-polar 60 M RTX 5MS Column

MS: Quadra pole detector with NIST Library

Software: GC MS Solution.

3.1 Chromatographic conditions

	Column:	Non-Polar 60 M RTX 5MS	
	Column Temperature:	750C for 2 min.	
	Flow rate:	1 ml/min.	
20	Injection volume:	1μl	
	Carrier Gas:	Helium (3ml/min)	
	Mode:	Split ratio.	
	IVIOUC.	Split radio.	

IV. RESULTS AND DISCUSSION: -

Mass spectra of methanolic extract of mangroves is shown in Table No. 4.1. While the mass fragments of unknown sample and Library standards are shown in Fig. 1 and 2. The mass spectra of mangrove bark extracts shows different peaks and the identification was made by retention time (R.T.) and National Institute of Standards and Technology (NIST) library search.

The mass spectra of Cynometra iripa show 4-Methylmannose, and 3-0- methyl-d-glucose in the methanol extract with percent area 62.16% and 37.84% respectively and their retention time are 28.211 and 27.317min. The compound like 9-12 Octadecadienoic acid (Linoleic acid), Ergosta- 8,24(28)-dien-3-ol,4,14-dimethyl and sitosterol with percent area, 58.22%, 19.48% and 14.48% respectively are abundantly found in methanolic extracts of Lumnitzera racemosa.

Many reports of antimicrobial properties are reported with isothiocyanates (Dornberger et al., 1975 and Iwu et al., 1991), polyamines (Spermidine) (Flayeh and Sulayman, 1987), Thiosulfinates (Tada et al., 1988) and glycosides (Murakami et al., 1993 and Rucker et al., 1992). Polyacetylenes deserve special mention. Form Bapleurum salicifolium Estevez- Braun et al., (1994) extracted a C₁₇ polyacetylene compounds, native of the Canory Islands. The compounds named as 8s-heptadeca-2(z), 9(z) diene-4, 6diyne-1, 8-diol, shows their inhibitory effects over bacteria like, S. aureas and B. subtilis but found inert effects over Gram positive bacteria as well as the fungus, Yeasts. Phytochemical studies of Abeysinghe and Wanigatunge (2006) proved that the mature leaf extract of A. marina shows the presence of bioactive compounds like, alkaloids, flavonoids, steroids and triterpenoids. phytochemical investigation of Rhizophora stylosa Griff. (Rhizophoraceae) reported one acetylated flavanol,3,7,0-diacetyl (-) eplcatechin (Anjuaneyulu et al., 2002). Gupta et al. (1980) extracted from the stem 5,7dihydroxyflavanone 4'-02-1 rhamnopyranosyl-β-D-glucopyranoside (I). The stem contains β-sitosterol, lupeol, Kempferol-3-glucoside and 5,7- dimethyl ether 4' rhamnoglucoside. The stem bark of B. variegata yields four substances viz. hentriacontane, octacosanol, β-sitosterol and sigmasterol (Anandaprakash, 1978). The known compounds from Rhizophora stylosa are 1,2,4-8 were characterized as (-) epicatechin (Lin et al., 2001b) (Foo et al., 1997) 3-0 acetyl(-) epicatechin (de Bruyne et al., 1999) (Ramesh et al., 2003) 3-3', 4' 5,7-0 pentacetyl (-) epicatechin (Laphookhieo et al., 2004) (Wan and Chan, 2004), (+) - afzelechin (Melchor et al., 2001 and Foo et al., 1997) Cinchanain Ib (Marerro et al., 2006) and proanthocyanidin B2 (Berenguer et al., 2006) (Foo et al., 1997), respectively, based on the detailed comparison of the H- and C-NMR spectral data with those of the literature reports (Li et al., 2007). Chifu et al. (2010) reported that esters of fatty acids have antimicrobial activity against oral pathogens like Streptococcus mutans, Candida albicans, Aggregatibacter actinomycetemcomitans, Fusobacterium nucleatum, and Porphyromonas gingivalis. Reports of Dewi et al. (2020) are also same of the earlier that fatty acid ethyl esters like linoleic acid and α-linolenic acid exhibited strong antibacterial activities against S. aureus and B. subtilis.

The presence of 9-12 Octadecadienoic acid / Linoleic acid present in the stem bark of Lunnitzeara racemose may be responsible for antimicrobial activity. While the observations of Matthew et al. (2019) shows a strong anti-inflammatory and antibacterial activity due to presence of linoleic acid while results of Stella et al. (2010) reported that presence of Sitosterol shows anti-inflammatory activity. The useful chemicals like 9-12 Octadecadienoic acid / Linoleic acid and Sitosterol are present in Lumnitzera racemose than Cynometra iripa. So, it may be having more antimicrobial activity in them. Further this study progresses towards antimicrobial and anti-inflammatory study.

Phytochemical analysis of the methanolic extracts of mangroves revealed that the presence of flavonoid, tannin, phenols, terpenoids and saponin derivatives. The antimicrobial properties of these secondary metabolites and from other extracts have also been reported. The presence of these compounds in the methanolic extracts could be the antimicribialy and anti-inflammatory active phytochemicals. Further study progresses over antimicrobial and anti-inflammatory activities of Lumnitzera racemose is essential.

Table No. 4. 1 Relative percentage composition of plant extracts of mangroves.

Sr. No.	Constituents	Retention Time (R. T.) (Min.)	Area %	Activity
Cynometr	ra iripa.			
1	4-Methylmannose	28.211	62.16	Unknown
2	3-O-Methyl-d-glucose	27.317	37.84	Unknown
Lumnitze	ra racemosa.		akta.	
1	9-12 Octadecadienoic acid / Linoleic acid	33.307	58.22	Antibacterial activity (Venkatesalu et al., 2004; McGaw et al., 2002)
2	Ergosta-8,24(28)-dien-3-ol,4,14- Dimethyl	34.101	19.48	Unknown
3	Sitosterol	30.717	14.48	Anti-inflammatory activity (Stella et al., 2010.)

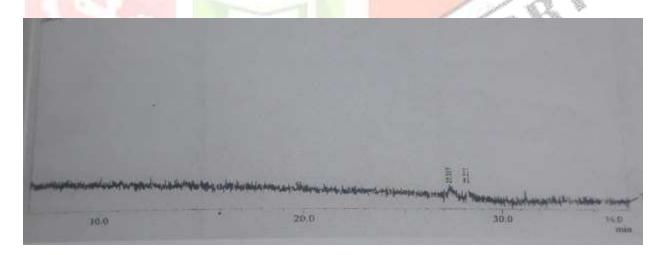


Figure 1. GCMS analysis of Cynometra iripa

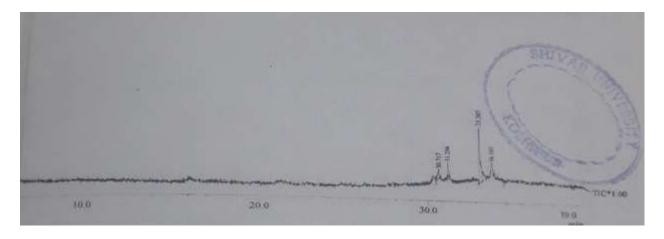


Figure 2. GCMS analysis of Lumnitzera racemosa

Phytochemical analysis of the methanolic bark extracts of mangroves revealed the presence of flavonoid, tannin, phenols, terpenoids and saponin derivatives. The antimicrobial properties of these secondary metabolites and from other extracts have also been reported. The presence of these compounds in the methanolic extracts could be the antifungally and anti-inflammatory active phytochemicals. The bark extract of Lumnitzera racemosa may be having more active phytochemicals with antimicrobial and anti-inflammatory activities due to presence of linoleic acid and sitosterol than the *Cynometra iripa*.

V. ACKNOWLEDGE: -

One of the authors is thankful to the research committee, Head Department of Botany and Principal, Dada Patil Mahavidyalaya, Karjat, for providing financial support to the present investigation.

REFERENCES:

- [1] Abeysinghe, P. D. and Wanigatunge, R. P. (2006). Evaluation of antibacterial activity of different mangrove plant extracts. Ruhuna Journal of Science, 1:104-112.
- [2] Anandaprakash, R. L. (1978). Natural constituents of Bauhinia variegata. Journal of Research into Indian Medicine, Yoga and Homeopathy **13**: 96-97.
- [3] Anjaneyulu, A. S. R., Anjaneyulu, V. and Rao, V. L. (2002). New Beyerane and Isopimarane Diterpenoids from *Rhizophora mucronata*, J. Asian Nat. Prod. Res., 4: 53-61.
- [4] Bandaranayake, W. M. (2002). Bioactivities, bioactive compounds and chemical constituents of mangrove plants. Wetland Ecol. Manage., 10: 421-452.
- [5] Berenguer, B., Sanchez, L. M., Quilez, A., Lopez-Barreiro, M., de Haro, O., Galvez, J., Martin, M. J. (2006). Protective and Antioxidant effects of Rhizophora mangle. L. Against NSAID-induced gastric ulcers. J. Ethnopharmacoal., 103: 194-200.
- [6] Chifu B. H.; Brian G. and Jeffery L. E. (2010). Antimicrobial activity of n-6, n-7 and n-9 fatty acids and their esters for oral microorganisms. Arch Oral Biol. 2010 Aug; 55(8): 555-560.
- [7] de Bruyne, T; Pieters, L., Declstra, H. and Vlietick, A. (1999). Condensed vegetable tannins: Biodiversity in structure and Biological activities. Biochem. Syst. Ecol. 27: 445-449.
- [8] Dewi K.; Misaki W.; Mai M.; Xiaonan X. and Kabuyama Y. (2020). Isamu Maeda Linoleic Acid, α-linolenic Acid, and Monolinolenins as Antibacterial Substances in the Heat-Processed Soybean Fermented With Rhizopus oligosporus. Biosci Biotechnol Biochem 84(6):1285-1290.
- [9] Dornberger, K; Bockel, V., Heyer, J., Schonfeld, C., Tonew, M. and Tonew, E. (1975). Studies on the isothiocyanates erysolin and sulforaphan and from Cardaria draba. Pharmaize, 30: 792-796.
- [10] Estevez-Braun, A., Estevez-Reyes, R., Moujir, L. M.; Ravelo, A.G. and Gonzalez, A.G. (1994). Antibiotic activity and absolute configuration of 8S-heptadeca-2(Z),9(Z)-diene-4,6-diyne-1,8-diol from Bupleurum salicifolium. J.Nat. Prod. 57: 1178-1182.
- [11] Flayeh, K. A and Sulayman, K. D. (1987). Antimicrobial activity of the amine fraction of cucumber (Cucumis sativus) extract. J. Appl. Microbiol., 3: 275-278.
- [12] Foo, L. Y., Lu, Y., Mcnabb, W. C., Waghon, G. and Ulyatt, M. J. (1997). proanthocyanidines from Lotus pendunculatus. *Phytochemistry*, **45**: 1689-1696.
- [13] Gupta, A. K., Vidyapati, T. J., Chauhan, J. S. (1980). Chemical examination of *Bauhinia variegate*. *Phytochemistry*, **64**:879-882.
- [14] Iwu, M. M., Unaeze, N. C., Okunji, C.O., Corely, D. G., Sanson, D. R. and Tempesta, M. S. (1991). Antimicrobial aromatic isothiocyanates from the essential oil of *Hippocratea welwitschii* roots. *Int. J. Pharmacogen.*, **29**: 154-158.
- [15] Kathiresan, K. and Bingham, B. L. (2001). Biology of mangroves and mangrove ecosystem. Adv. Mar. Biol., 40: 81-251.
- [16] Kokpol, U.; Miles, D. H.; Payne, A. M. and Chittawong, V. (1990). Chemical constituents and bioactive compounds from mangrove plants. In: Att-ur- Rahman (ed), Studies in Natural Products Chemistry, Elsevier Science Publishers B. V. Amsterdam. Pp. 7: 175-195.
- [17] Laphookhieo, S., Karalai, C. and Ponglimanont, C. (2004). New Sesquiterpenoid and Triterpenoids from the Fruits of Rhizophora mucronata. Chem. Pharm. Bull., 52: 883-885.

- [18] Li, D. L., Li, X. M. Peng Z. Y. and Wang, B. G. (2007). Flavanol derivatives from *Rhizophora stylosa* and their DPPH radical scavenging activity. *Molecules*, 12: 1163-1169.
- [19] Lin, J. K., Tsai, S. H. and Lin-Shiau, S.Y. (2001a). Antiinflammatory and antitumor effects of flavonoids. *Drugs Future*, 26: 145-157.
- [20] Marrero JA, Feng Z, Wang Y, Nguyen MH, Befeler AS, Roberts LR, et al. Alpha-fetoprotein, des-gamma carboxyprothrombin, and lectin-bound alpha-fetoprotein in early hepatocellular carcinoma. *Gastroenterology* 2009; **137**: 110-118
- [21] Matthew, J. K.; Srihari, K.; Tina, C.; Huijing, W.; Clare M.; Lena, O.; Magnus H.; Dionicio, S. and Alan S.(2019). Linoleic Acid Esters of Hydroxy Linoleic Acids Are Anti-Inflammatory Lipids Found in Plants and Mammals. *J Biol Chem*, **294**(27):10698-10707.
- [22] McGaw, L.J.; Jäger, A.K.; Van Staden, J. (2002). Isolation of antibacterial fatty acids from Schotia brachypetala. Fitoter., 73: 431-433.
- [23] Melchor, G., Armenteros, M., Fernandez, O., Linares, E. and Fragas, I. (2001). Antibacterial activity of *Rhizophora mangle* bark. *Fitoterapia*, 17:689-691.
- [24] Murakami, A., Ohigashi, H., Tanaka, S., Tatematsu, A. and Koshimizu, K. (1993). Bitter cyanoglucosides form *Lophira alata*. *Phytochemistry*, 32: 1461-1466.
- [25] Ramesh, C., Mahender, G., Ravindranath, N; Das and Mild, B. A (2003). Highly selective and remarkably easy procedure for deprotection of aromatic acetates using Ammonium Acetate as a natural catalyst in aqueous medium. Terahedron, 59: 1049-1054.
- [26] Rucker, G., Keherbanum, S., Sakulas, H., Lawong, B. and Goeltenboth, F. (1992). Acetylenic glucosides from *Microglossa pyrifolia*. *Planta Med.* 58: 266-269.
- [27] Saenger, P. (2002]. Mangrove Ecology, Silviculture and Conservation. Springer. ISBN 978-94-015-9962-7.
- [28] Stella L.; Ioannis L.; George P. C. and Paraskevi M. (2010). β-Sitosterol exhibits anti-inflammatory activity in human aortic endothelial cells. *Molecular Nutritional*: **54**(4): 551-558.
- [29] Tada, M., Hiroe, Y., Kiyohara, S. and Suzuki, S. (1988). Nematicidal and antimicrobial constituents from *Allium grayi* Regel and *Allium fistulosum* L.var. caespitosum. *Agric. Biol. Chem.*, 52: 2383-2385.
- [30] Venkatesalu, V.; Sundaramoorthy, P.; Anantharaj, M.; Gopalakrishnan, M.; Chandrasekaran, M. (2004). Studies on the fatty acid composition of marine algae of Rameswaram coast. Seaweed Res. Util., 26: 83-86.
- [31] Wan, S. B and Chan, T. H. (2004). Enantioselective of Afzelechin and Epiafze lechin. Tetrahedron, 60: 8207-8211. www.nhmi.org/mangroves.

